# Novel Approaches and Technologies to Assess Genotoxic Modes of Action

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HESI GTTS Workshop, March 22 + 23, 2018, Potsdam, Germany



#### Agenda

- Introduction
- MoA via genomic biomarkers
  - Approach taken
  - Transcriptomics
    - Tgx-DDI biomarker (DNA Damage Inducing)
    - Connectivity mapping
  - ToxTracker
- Conclusions, part 1
- Pyrrozolidinly alkaloids use of MoA to support potency ranking
- Conclusions, part 2 and open questions



#### Introduction

- P&G has a goal to develop/establish a reliable & predictive in vitro method for identifying mode of action (MoA)
- Initial interest is in disseminating primary (direct) from secondary (indirect) effects for its impact on risk assessment
- Approach taken is to compare different methodologies using genomic biomarkers that have been developed for classifying chemicals by MoA
  - Insights into what drives the toxic response
  - Draw conclusions regarding primary/secondary genotoxicity
  - Ultimate goal goes beyond classification

connect to data rich chemicals: read across

- Many different ways to approach MoA, other examples in WS
- Can support each other and reduce uncertainty

High burden of proof for regulatory decision making



#### Approach

- Compare different methodologies that have been developed for classifying chemicals by MoA → TGx-DDI, C-Map, ToxTracker
- Examine the impact of the genomic platform used, and reduction of information
  - Test 22 chemicals using:
    - 'All-in-one approach' where samples from a relevant genotoxic endpoint (flow MN assay) are also used for genomic analysis (Affymetrix).
    - L1000 Expression Profiling (Peck et al. Genome Biology 2006). Uses "Landmark Genes" that reflect full genome expression profiles. (Cheaper, faster, more high throughput)
      - Analyze results and compare both using Connectivity Mapping (CMap)
    - Compare with results from coded testing with Toxtracker, a stem cell-based reporter assay



### **Selected Chemicals**



### **Combination Approach Overview**





1) Micronuclei in Tk6 cells



# False Positive



#### **True Positive**



### **Dose selection for gene analysis**

- For Affy
  - Cytotoxicity
  - MN response
  - Total no of affected genes

#### For CMap

- Selected single dose from Affy data
- Moderate level of induction



### L1000 Expression Profiling Overview

- TK6 cells were treated for 4 hours, then processed (crude lysates) and frozen at -80C.
- Selected 1-4 doses per compound using results from combination approach.
- 4 independent, randomized 96-well experiments performed.
- Transferred to a 384 well plate and sent to Genometry for analysis.



# Data analysis 1 - TGx-DDI

- Result of a HESI Toxicogenomics team project
- Identification of DNA Damage Inducing (DDI) agents (no anuegens)
- Dose optimization protocol using qRT- PCR of stress response genes (CDKN1A; GADD45A; ATF3)
- Followed by microarray (Agilent) analysis
- 65-gene 'DDI' signature was determined

Our data were analyzed by Health Canada (Andrew Williams, Carole Yauk)

Environmental and Molecular Mutagenesis 56:505-519 (2015)

Development of a Toxicogenomics Signature for Genotoxicity Using a Dose-Optimization and Informatics Strategy in Human Cells

Heng-Hong Li,<sup>1,2</sup> Daniel R. Hyduke,<sup>1,2,3</sup> Renxiang Chen,<sup>1,2</sup> Pamela Heard,<sup>4</sup> Carole L. Yauk,<sup>5</sup> Jiri Aubrecht,<sup>4</sup> and Albert J. Fornace Jr.<sup>1,2,6</sup>\*

# TGx-DDI – 65 gene set



# Data analysis 2: Whole genome information The Connectivity Mapping (CMap) Concept



#### CMap Analysis: Utilization of <u>3 doses</u> – Affy vs L1000



#### Affymetrix ~ 40,000 genes



True Positive (11)
True Negative (5)
False Positive (6)



#### L1000 Expression Profiling ~ 1,000 genes

# CMap Analysis: Use of <u>one</u> target dose - Impact of platform



#### L1000 Expression Profiling ~ 1,000 genes





## Use of reporter genes – ToxTracker assay

#### Overview

- Uses 6 mES GFP reporter cell lines
- High sensitivity and specificity, according to ToxTracker internal validation
- International validation effort ongoing
- Mechanistic insight into toxicity

| Biological damage | Biomarkers   |  |  |  |
|-------------------|--------------|--|--|--|
| DNA damage        | Bscl2, Rtkn  |  |  |  |
| Oxidative stress  | Srxn1, Blvrb |  |  |  |
| Protein damage    | Ddit3        |  |  |  |
| Cellular stress   | Btg2         |  |  |  |



### Data analysis 3: ToxTracker

|  |                    |   | DNA d | DNA damage |     | Oxidative stress |     | UPR |     | p53 |  |
|--|--------------------|---|-------|------------|-----|------------------|-----|-----|-----|-----|--|
| $ \rightarrow 1001 \text{ KAU} $   |                    |   | -S9   | +S9        | -S9 | +S9              | -S9 | +S9 | -S9 | +S9 |  |
|  | KED <sub>0</sub>   | Test compounds  |       |            |     |                  |     |     |     |     |  |
| Overall results<br>after decoding<br>22 chemicals  | TN                 | Cyclohexanone<br>Mannitol<br>Amitrol<br>2-Deoxy-D-glucose<br>Sodium diclofenac  |       |            |     |                  |     |     |     |     |  |
|  | FP                 | Ethionamide<br>Sodium saccharin<br>Tertiary-butylhydroquinone<br>Curcumin<br>Eugenol<br>Quercetin   |       |            |     |                  |     |     |     |     |  |
|  | ТР                 | 5-Fluorouracil<br>Etoposide<br>Vinblastine<br>Camptothecin<br>Methyl methanesulfonate<br>Hydrogen peroxide<br>Hydroquinone<br>ENU<br>O-toluidine<br>Colchicine<br>Sodium arsenite |       |            |     |                  |     |     |     |     |  |
| Positive (>2-fold induction)<br>Weak positive (1.5 to 2-fold<br>Negative (<1.5-fold inductio<br>Inconclusive results | l induction)<br>n) | Controls<br>Cisplatin<br>Diethyl maleate<br>Tunicamycin<br>Aflatoxin B1   |       |            |     |                  |     |     |     |     |  |

Looking at cutoff by fold increase only (yes/no) misses important information

# Data analysis 3: ToxTracker

### Example: tert-Butylhydroquinone



#### **Conclusions part 1**

- All three methods show good predictive capacity for set of 22 coded compounds
- C-map and ToxTracker can reveal MoA insights
- C-map shows promise for 'grouping' of chemicals since it takes into account toxicological signatures across pathways
- Increasing trend to "condense" (~40,000 genes → 1000) leads to information loss
- Next steps: More in depth analysis of one specific MoA (oxidative stress), added additional chemicals
- Described methods can inform MoA and therefore help risk assessment



#### **Pyrrozolidinly alkaloids**

- Pyrrolizidine Alkaloids (PA) are constituents of certain plant families (defense mechanism)
- There are hundreds of PAs but 1,2-unsaturated PAs mainly relevant for safety assessment
- MoA understood/supported well, via in vivo genotoxicity and carcinogenicity data
  - Direct acting mutagen needing metabolic activation
  - Strongly hepatotoxic (poisoning of feedstock, human cases)
- Exposure limits were suggested for PAs in Europe [ECHA 2017 limit: 0.07 mg/kg bw/day]
- Applies for all PA's [sum], but value is <u>driven by the most potent PA</u>
- Relative potencies seem to strongly vary, as a consequence of structural differences [Merz and Schrenk. 2016. Toxicology Letters 263. p44–57]

Can MoA information be used to 'group' PAs, and can 'Key Events' be used to derive relative potency factors (RPF)?





#### Pyrrozolidinly alkaloids

Comprehensive testing program ongoing, aiming to build convincing case based on in vitro and modeling data:

- Cancer potency depends on reactivity of 1,2-unsaturated PAs
  - 1. Measure rate of reactive metabolite formed (liver microsomes, primary liver cell culture, HepaRG)
  - 2. Measure their relative genotoxicity potency in metabolically component cell system (micronuclei in HepaRG)
  - 3. Link 1. and 2. via DNA adduct formation rates.
  - 4. Consider key additional factors (e.g, oral absorption, fate of 'Noxides', cellular uptake)
  - 5. Use 1-4 to calculate RPFs that enable data-driven risk assessment for PAs

Linking expected toxicity via common MoA is prerequisite for acceptance of RPFs



#### **Relative Potency of PAs, initial results**

- Developed flow-based micronucleus assay using HepaRG cells (support/training by ILS)
- HepaRG express key enzymes responsible for activation of PAs, e.g, Cyp 3A4
- Investigated 18 PAs to date
- Dose-response curves generated for all (single replicates), aiming to establish optimum dose-range for main studies
- Main studies in triplicates: tight dose-spacing to enable BMD modeling
- Examples:
  - Lasicarpine (suggested RPF 1.0)
  - Lasicarpine N-oxide (RPF ?)
  - Heliotrine (suggested RPF 0.3)
- Critical Effect Size chosen for modeling: 2- fold increase over background



# Lasiocarpine, RPF 1.0



Calculated with PROAST BMD software

# Lasiocarpine, RPF 1.0



# Heliotrine, RPF 0.3



Potency compared to lasiocarpine: 0.1

## Lasiocarpine N Oxide – RPF ?



#### **Conclusions 2**

- Pyrrolizidine Alkaloids (PA) are plant-based impurities with defined genotoxic and carcinogenic MoA
- Current risk management measures define toxicity thresholds that base on assumption all PAs are equipotent
- MIE and KE's can be used to delineate potency
- Additional modulating factors need to be defined per PA for proper calculation of RPF (e.g, oral and cellular uptake, gut metabolism)
- Initial data from micronucleus testing in HepaRG cells show strong differences in potency across PAs
- We believe all parameters can be modeled via *in vitro* and *in silico* data



#### **Open questions**

- MOA/AOPs how high is the burden of proof?
- How can regulatory acceptance be supported?
- How to address 'uncertainty' in this context?
- Will the genetox community engage to help drive developments of AOP?



# Acknowledgements

Procter & Gamble Stefan Pfuhler Tom Downs Nadira DeAbrew Raghu Kainkaryam Yuching Shan Rachel Adams Genometry Justin Lamb Willis Read-Button

Health Canada Andrew Williams Carol Yauk <u>Toxys</u> Giehl Hendrichs Paula von Rossum





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